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Reliability of ultrasound hepatorenal index and magnetic resonance imaging proton density fat fraction techniques in the diagnosis of hepatic steatosis, with magnetic resonance spectroscopy as the reference standard

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27

28 **Abstract**

29 **Purpose:** To evaluate the reliability of ultrasound hepatorenal index (US-
30 HRI) and magnetic resonance imaging proton density fat fraction (MRI-
31 PDFF) techniques in the diagnosis of hepatic steatosis, with magnetic
32 resonance spectroscopy proton density fat fraction (MRS-PDFF) as the
33 reference standard.

34 **Materials and Methods:** Fifty-two adult volunteers (30 men, 22 women;
35 age, 31.5 ± 6.5 years) who had no history of kidney disease or
36 viral/alcoholic hepatitis were recruited to undergo abdominal US, MRI,
37 and MRS examinations. US-HRI was calculated from the average of
38 three pairs of regions of interest (ROIs) measurements placed in the liver
39 parenchyma and right renal cortex. On MRI, the six-point Dixon

40 technique was employed for calculating proton density fat fraction (MRI-
41 PDFF). An MRS sequence with a typical voxel size of 27 ml was chosen
42 to estimate MRS-PDFF as the gold standard. The data were evaluated
43 using Pearson's correlation coefficient and receiver operating
44 characteristic (ROC) curves.

45 **Results:** The Pearson correlation coefficients of US-HRI and MRI-PDFF
46 with MRS-PDFF were 0.38 ($p=0.005$) and 0.95 ($p<0.001$), respectively. If
47 MRS-PDFF $\geq 5.56\%$ was defined as the gold standard of fatty liver
48 disease, the areas under the curve (AUCs), cut-off values, sensitivities
49 and specificities of US-HRI and MRI-PDFF were 0.74, 1.54, 50%, 91.7%
50 and 0.99, 2.75%, 100%, 88.9%, respectively. The intraclass correlation
51 coefficients (ICCs) of US-HRI and MRI-PDFF were 0.70 and 0.85.

52 **Conclusion:** MRI-PDFF was more reliable than US-HRI in diagnosing
53 hepatic steatosis.

54

55 **Introduction**

56 Nonalcoholic fatty liver disease (NAFLD) is the most common liver
57 disorder. A meta-analysis reported a prevalence of 24% in the worldwide
58 population [1]. NAFLD is also considered an important cause of fibrosis
59 progression, nonalcoholic steatohepatitis (NASH), and hepatocellular
60 carcinoma (HCC) [2]. Based on the literature, NAFLD has shown a

61 strong association with coronary artery disease, osteoporosis, metabolic
62 syndrome [3], and rheumatoid arthritis [4]. The prevalence of NAFLD
63 varies with age, gender, and weight status [5]. Early detection and
64 quantification of hepatic steatosis play an important role in treatment
65 because NAFLD can be treated by control of diabetes, weight loss or
66 lifestyle modification [6].

67 Liver biopsy is still described as the reference standard for
68 quantifying liver fat content [7]. However, liver biopsy is invasive, with
69 risk of bleeding and other miscellaneous complications, and also has
70 potential for sample bias and inter- and intra-observer variability [7]. A
71 noninvasive and robust method of hepatic steatosis measurement is
72 necessary not only for early detection of hepatic steatosis, but also for
73 monitoring during treatment. Ultrasound (US) is a widely used
74 noninvasive method of assessing fatty liver disease, particularly as a
75 screening tool, because of its low cost, safety and accessibility. The ratio
76 between echogenicity of the liver tissue and renal cortex, called the
77 hepatorenal index (US-HRI), has been commonly used to estimate the
78 degree of steatosis. This ratio is positively correlated with the fat
79 percentage [8]. However, US-HRI has limitations such as variation of
80 HRI values among machines and operators.

81 MR scanners provide additional noninvasive alternatives for
82 hepatic steatosis measurements by directly quantifying fat content

83 fraction based on the difference in resonance frequencies between water
84 protons and fat protons. Non-invasive magnetic resonance spectroscopy
85 (MRS) providing proton density fat fraction (MRS-PDFF) has been
86 considered an alternative method for evaluating liver fat content. This
87 method seems to be reasonable and is a potential alternative for
88 quantifying liver fat, given that it has actually been shown to be very
89 accurate in comparison to histological diagnosis [9]. However,
90 performing this technique requires the addition of a special software
91 package usually not available by default. It is also a time-consuming
92 technique, which also hinders its widespread use.

93 Magnetic resonance imaging proton density fat fraction (MRI-
94 PDFF) is a newer technique proposed in the diagnosis of hepatic
95 steatosis. This technique can be considered a hybrid methodology, as it
96 combines the advantages of complex-based fitting and magnitude-based
97 fitting techniques to estimate fat fraction. The multi-echo adaptive fitting
98 technique uses the Levenberg-Marquardt fitting algorithm to solve for the
99 values of water and fat signal intensity. A multi-step nonlinear fitting
100 procedure is then performed to adaptively update the fat and water
101 signal fractions based on magnitude signal equations with a multi-peak
102 fat spectral model [10]. The major advantages of this technique over
103 MRS are that (1) it is technically easier to implement, (2) the software
104 package needed is commonly available on conventional MRI units, and

105 (3) the examination time is short (less than 15 minutes). We suspect this
106 technique to be a potential replacement for US, or even a first line tool
107 for diagnosis and management of hepatic steatosis.

108 The purpose of this study was to evaluate the reliability of fat
109 quantification by US (US-HRI) and MRI-PDFF techniques in the
110 diagnosis of hepatic steatosis, with MRS-PDFF as the reference
111 standard.

112

113 **Materials and Methods**

114 **Subjects**

115 Adult volunteers having no known hepatic nor renal disease were
116 randomly recruited over a period of 19 months (Nov. 2018 - Apr. 2020)
117 including students and staff on a certain campus. There was no one
118 outside this location. Participation was strictly voluntary, and participants
119 did not receive any money. In Japan, annual health check-up for all
120 employees and students are required by law, and the data, including
121 serum alanine aminotransferase (ALT), aspartate aminotransferase
122 (AST), lactate dehydrogenase (LDH), total bilirubin and serum creatinine,
123 were used to rule out liver and kidney disease. Medical school staff and
124 students are tested for HBV and HCV at the same time. All of these
125 costs are paid by the university.

126 Subjects with known diabetes mellitus, hepatitis B or C virus
127 infection, excess alcohol intake (> 20g/day), thyroid disease, and long-
128 term drug therapy such as corticosteroids were excluded from this study.
129 US and MRI examinations were performed on the same day.

130 This prospective study was approved by the research ethics
131 committee of our institutional review board (Gunma University Graduate
132 School of Medicine, Japan), and written informed consent was obtained
133 from all participants. There were no relevant conflicts of interest.

134

135 **US-HRI**

136 US examination was performed using a HI VISION Ascendus
137 (Hitachi Ltd, Tokyo, Japan) unit equipped with a curved phased-array
138 probe EUP-C715 (1-5 MHz). Imaging examinations and measurements
139 were performed by a board-certificated diagnostic radiologist (ATT) with
140 twenty years of experience. Instrument settings such as gain and depth
141 were adjusted by the operator, depending on the body size of
142 participants.

143 An image with the liver and right kidney in the same field of view
144 was obtained in the left lateral decubitus position from the right sagittal or
145 right intercostal approach. Regions of interest (ROIs) with a size of 100
146 mm² in the liver parenchyma and 25 mm² in the right renal cortex were

147 selected (Fig 1). The ROIs were selected to avoid blood vessels and
148 situated near the center of the image to be the same depth, gain, and
149 mean gray-scale of the pixels [8]. If images of the liver and right kidney
150 could not be obtained in the same field of view in the left lateral
151 decubitus position, the liver and right kidney were imaged in the prone
152 position from a right sagittal approach. US-HRI was calculated as the
153 ratio of the echogenicity of the hepatic parenchyma to the echogenicity
154 of the right renal cortex. This procedure was repeated five times with two
155 ROIs on each scan. The mean of the three closest values was used with
156 the difference between the values obtained being less than 0.2 [8].

157

158 **Fig 1. HRI measurement on a volunteer with mild hepatic steatosis**
159 **(HRI = 2.33).**

160

161 **MRI-PDFF**

162 The six-point Dixon technique was employed using modeling of a
163 multi-echo adaptive fitting approach (LiverLab, Siemens Medical
164 Systems, Erlangen, Germany) with a 3.0-Tesla magnet (MAGNETOM
165 Skyra, Siemens Medical Systems, Erlangen, Germany). Multi-axial
166 images were obtained by the three-dimensional gradient-recalled-echo
167 (3D-GRE) pulse sequence with a 24-channel spine matrix coil and 18-

168 channel body matrix coil. To estimate water and fat signals, six echoes
169 with whole liver coverage were conducted in a single breath-hold (12
170 seconds). A short TR (9 ms) and a small flip angle ($\alpha = 40$) were used in
171 this pulse sequence with the aim to minimize T1 bias and T2*-effect.
172 Other imaging parameters were: field of view (FOV) 350 mm, matrix 95 x
173 160, slice thickness 3.5 mm, echo time (TE) 1.12, 2.46, 3.69, 4.92, 6.15,
174 7.38 ms, parallel imaging factor of 2 x 2, and spatial resolution of 2 x 2 x
175 2 mm³. The Dixon sequence automatically generated series of water, fat,
176 water percentage, fat percentage, goodness-of-fit, R2* map, T2* map,
177 and fat fraction. ROIs were manually set in the right hepatic lobe to be as
178 large as possible while avoiding margins, biliary tract, gallbladder,
179 artifact, and large vessels. The goodness-of-fit was an indication of fitting
180 residual errors of the fat percentage result, and MRI-PDFF were
181 calculated as shown on Fig 2. The MR imaging and measurements were
182 performed by a technologist (BVT) with 8 years of experience in MRI.

183

184 **Fig 2. MRI-PDFF measurement on a volunteer with mild hepatic**
185 **steatosis (MRI-PDFF = 8.3%).**

186

187 **MRS-PDFF**

188 Immediately after MRI-PDFF measurements, a single-voxel MRS
189 was performed to measure fat content as the reference standard. A high-
190 speed T2-corrected multi-echo (HISTO) sequence was employed with a
191 15 seconds breath-hold. A stimulated echo acquisition mode (STEAM)
192 was applied with the following parameters: voxel size of 30 mm x 30 mm
193 x 30 mm (27 ml), TR of 3000 ms, 5 spectra at TE of 12, 24, 36, 48 and
194 72 ms, number of excitation (NEX) 1, and receiver bandwidth of 1200
195 Hz/Px. A voxel was placed in a homogeneous portion of the liver
196 avoiding margins, biliary tract, gallbladder, artifact, and large vessels. On
197 MRS, with the axial image active, the scroll nearest tool was used to
198 select the coronal and sagittal image to the voxel position on the axial
199 much the same as normal spectroscopy positioning.

200 Data were baseline corrected, phase-corrected, averaged and
201 Fourier transformed. Levenberg-Marquardt curve fitting was performed
202 using a combined Lorentzian-Gaussian model to calculate the area
203 under the curve of fat and water peaks [11,12]. MRS-PDFF was
204 calculated as shown on Fig 3. The color bar map showed the amount of
205 fat as a percentage. The measurement of MRS was performed by one
206 technologist (KU) with 15 years of experience.

207

208 **Fig 3. MRS-PDFF measurement on a volunteer with mild hepatic**
209 **steatosis (MRS-PDFF = 10%).**

210

211 Measurements of US-HRI, MRI-PDFF and MRS-PDFF were
212 separately performed without knowledge of the results of other
213 measurements. However, as much as possible, the ROI for MRI-PDFF
214 measurement was placed in the same position as the voxel in MRS-
215 PDFF measurement in the right hepatic lobe, since the liver fat
216 distribution may be inhomogeneous, potentially affecting the signal
217 intensity. According to Szczepaniak and colleagues [13], grade 0
218 (normal), grade 1 (mild), grade 2 (moderate), and grade 3 (severe) were
219 defined as corresponding to 0 - $\leq 5.56\%$, 5.56% - $\leq 10\%$, 10% - $\leq 20\%$,
220 and $>20\%$ fat content, respectively. For MRS-PDFF, 5.56% fat was
221 considered the cut-off value for this study.

222

223 **Statistical techniques**

224 Pearson's correlation was used to correlate the US-HRI and MRI-
225 PDFF with MRS-PDFF. Receiver operating characteristic (ROC) curves
226 including the area under curves (AUC) values were calculated to
227 evaluate the accuracy of US-HRI and MRI-PDFF in determining hepatic
228 steatosis. Optimal cut-off values giving sensitivity and specificity were
229 computed by using Youden index.

230 The reproducibility of US-MRI and MRI-PDFF measurements was
231 evaluated in 15 randomly selected subjects, who underwent two
232 repeated measurements within an interval of 100 days to avoid the
233 alteration of hepatic fat content over time [14]. Limits of agreement using
234 the mean value of the two different measurements were calculated
235 according to Bland-Altman analysis [15].

236 All analyses were conducted using the statistical software SPSS
237 version 25.0 (SPSS Inc. Chicago, IL), and $p < 0.05$ were considered
238 significant.

239

240 **Results**

241 **Participants**

242 A total of 52 participants (age, 31.5 ± 6.5 years [mean \pm SD];
243 range, 20 to 50) matched the inclusion and exclusion criteria, with 30
244 men (25-50 years) and 22 women (20-35) included in this study. Body
245 mass index (BMI) was $23.12 (\pm 3.62 \text{ kg/m}^2)$ according to the WHO
246 formula.

247

248 **Diagnosis of hepatic steatosis based on MRS-PDFF**

249 MRS-PDFF ranged from 1.0 to 16.7% ($5.3\pm 3.9\%$ [mean \pm SD]).
250 When the cut-off value was 5.56% on MRS-PDFF for the diagnosis of
251 hepatic steatosis, sixteen subjects (30.8%) had mild to moderate hepatic
252 steatosis. There were no subjects with severe hepatic steatosis.

253

254 **Correlations of US-HRI and MRI-PDFF with MRS-PDFF**

255 US-HRI ranged from 0.95 to 2.33 (1.4 ± 0.3). The Pearson
256 correlation coefficient between US-HRI and MRS-PDFF was significant
257 but weak ($r=0.38$, $p=0.005$; Fig 4). MRI-PDFF ranged from 0.2 to 15.4%
258 (3.8 ± 3.5). The Pearson correlation coefficient between MRI-PDFF and
259 MRS-PDFF showed excellent linear correlation ($r=0.95$, $p<0.001$; Fig 5).

260

261 **Fig 4. Correlation between US-HRI and MRS-PDFF.**

262 **Fig 5. Correlation between MRI-PDFF and MRS-PDFF.**

263

264 **Diagnostic accuracy**

265 With a 5.56% cut-off for MRS-PDFF, there were 16/52 participants
266 who had mild to moderate steatosis. MRI-PDFF showed higher
267 sensitivity (100%) and similar specificity (88.9%), compared to US-HRI
268 (50% and 91.7%, respectively) for the diagnosis of hepatic steatosis. The

269 cut-off values for MRI-PDFF and US-HRI were 2.75% and 1.54,
270 respectively (Table 1). The AUC value of MRI-PDFF (0.99) was higher
271 than US-HRI (0.74) (Fig 6). On ultrasound, the quantity of accuracy
272 (ACC) was 78.85%. Meanwhile, the ACC of MRI-PDFF was 84.62%.

273

274

275 **Table 1. Diagnostic performance of US-HRI and MRI-PDFF.**

	MRI-PDFF	US-HRI
Sensitivity	1.0	0.5
Specificity	0.889	0.917
AUC	0.99 ($p<0.001$)	0.74 ($p=0.006$)
Cut-off point	2.75 (%)	1.54
Quantity of accuracy	84.62%	78.85%

276

277

278 **Fig 6. ROC curve using the reference of 5.56% as cut-off point in**
279 **defining diagnostic performance.**

280

281 **Reproducibility**

282 The correlation coefficients for two repeated measurements of US-
283 HRI and MRI-PDFF were 0.70 ($p < 0.001$) and 0.85 ($p < 0.001$),
284 respectively. Bland-Altman analysis showed an excellent agreement
285 between two measurements of MRI-PDFF with the mean of difference of
286 0.13 percentage points (pp) (limits of agreement [LOA], -1.99 pp and
287 2.25 pp). The mean of difference between two measurements of US-HRI
288 was 0.02 (LOA, 0.47 and 0.51) (Fig 7 and 8).

289

290 **Fig 7. Bland-Altman plots for variability of PDFF measurements**
291 **generated using MRI.**

292 The central line shows the mean of the differences between two PDFF
293 measurements; the dashed lines show upper (mean + 1.96 SD) and
294 lower (mean - 1.96 SD) limits of agreement. Here, the mean difference is
295 0.13 pp, while the limits of agreement are -1.99 pp and 2.25 pp,
296 indicating that 95% of the differences between these two measurements
297 are within this range. The width interval is 4.24 pp.

298 **Fig 8. Bland-Altman plots for variability of HRI measurements**
299 **generated using ultrasound.**

300 The central line shows the mean of the differences between two HRI
301 measurements; the dashed lines show upper (mean + 1.96 SD) and
302 lower (mean - 1.96 SD) limits of agreement. Here, the mean difference is

303 0.02, while the limits of agreement are -0.47 and 0.51, indicating that 95
304 % of the differences between these two measurements are within this
305 range. The width interval is 0.98.

306

307 **Discussion**

308 In the current study, we found that MRI-PDFF showed excellent
309 linear correlation with MRS-PDFF (the gold standard in this study), and
310 its sensitivity for the diagnosis of hepatic steatosis was 100%, while that
311 of US-HRI was 50%. The reproducibility of MRI-PDFF was also very
312 good with a mean difference between two measurements of only 0.13
313 pp. MRI-PDFF is technically easier to implement than MRS-PDFF. The
314 examination time is only 15 min., and does not require any special
315 software package. To our knowledge, this was the first study directly
316 comparing the reliability of MRI-PDFF and US-HRI in quantifying liver fat
317 content.

318 According to a study comparing US-HRI and MRS-PDFF in 121
319 volunteers [8], there was a very good correlation between the two
320 techniques ($r=0.89$, $p<0.001$), thus it was concluded that US was valid
321 enough for the identification, assessment and quantification of hepatic
322 steatosis. On the other hand, in another study comparing US-HRI and
323 MRI-PDFF in 34 overweight adolescents [16], there was only a moderate

324 correlation between the two ($r=0.487$, $p=0.003$), and that report
325 concluded that US can be used as a screening tool for non-alcoholic
326 fatty liver diseases, but the diagnosis should be confirmed with MRI-
327 PDFF. The disagreement between these findings is unexpected, since
328 both MRS-PDFF and MRI-PDFF are measurement methods that use the
329 difference in the resonance frequencies of water and lipid protons, and
330 there should, theoretically, be no significant difference between the two
331 measurements. However, while the two methods are based on the same
332 physical principle (the small difference in resonance frequency between
333 water molecule protons and fat molecule protons), actual signal
334 processing is not the same. The Dixon method acquires signal when the
335 water molecule protons and the fat molecule protons are in-phase and
336 when they are in opposed-phase. Then, the sum or difference of these
337 signals are calculated pixel-by-pixel. These signals are processed into
338 images, and a ROI is selected for measurement. The in-phase and
339 opposed-phase images are acquired by selecting different TEs on
340 gradient echo (GRE) sequences [17]. In theory, for a given TE, all
341 protons will be in-phase or in opposed-phase, but in reality, the local
342 molecular environment of the protons is not completely homogenous,
343 and this will, albeit slightly, alter the resonance frequency. In addition,
344 technical limitations of hardware and static field inhomogeneity limit the
345 accuracy of TE (ms) to about the second decimal point.

346 On the other hand, MRS measures water molecule protons and fat
347 molecule protons directly and separately for a given voxel. No images,
348 sums or differences are involved, and minor variations in frequency due
349 to the state of protons become part of the distribution of frequency when
350 graphed [18]. There is no spatial information, and a large voxel is
351 needed for sufficient signal-to-noise ratio, but generally speaking, it is the
352 most accurate method of measurement.

353 Additional factors, such as the variation of T1 relaxation time by
354 TR, make it near impossible to make data acquisition completely
355 identical while maintaining clinical feasibility. Given these factors, the
356 different values are not surprising, and the difference in the slopes of the
357 linear correlation graphs is also understandable.

358 These limitations notwithstanding, the results of the current study
359 indicated that MRI-PDFF and MRS-PDFF were interchangeable, and
360 MRI-PDFF, which is the simpler method, may be ideal in the clinical
361 setting.

362 In the current study, the optimum cut-off value of MRI-PDFF for
363 diagnosing fatty liver was 2.75%, with sensitivity and specificity of 100%
364 and 88.9%, respectively. These results were consistent with a study of
365 94 subjects in determining the accuracy of MRS-PDFF using
366 histopathologic analysis as the standard, showing sensitivity of 100%
367 and specificity of 79% [19]. In a study investigating the accuracy of MRI

368 in quantifying liver fat in 86 children, the authors found a slightly higher
369 optimum MRI-PDFF threshold value of 5.1% with a sensitivity and
370 specificity of 95% and 100%, respectively [20].

371 Since liver fat distribution may be inhomogeneous, the signal
372 intensity on MRI may also be inhomogeneous. To the best of our
373 knowledge, no imaging technique can adequately evaluate
374 inhomogeneous fat distribution. When evaluating therapeutic efficacy, if
375 fat distribution is not uniform, PDFF should be evaluated at exactly the
376 same region before and after treatment. This is simple to accomplish on
377 MRI because we can easily confirm the inhomogeneity of fat distribution
378 visually. Although MRS-PDFF is an accurate measurement method, the
379 inability to identify non-uniform fat distribution is a major drawback.

380 This study used US-HRI as a prevalent imaging technique for the
381 diagnosis of hepatic steatosis in a routine setting. However, the current
382 study showed only average agreement between two measurements, with
383 an intra-observer correlation coefficient of 0.70 ($p < 0.001$). Despite being
384 a popular method, US-HRI has shown diverse results. According to data
385 compiled and published by Chauhan and colleagues [21], threshold
386 values varied from 1.24 to 2.02, sensitivity from 62.5% to 100%, and
387 specificity from 54% to 96%. A common cause mentioned for the wide
388 variation in ultrasound was its greater sensitivity to larger proportions of

389 fat. Machine and operator dependence also commonly contributed to the
390 wide variation of results [22].

391 The incomparability of US-HRI measurements from different
392 machines or different operators limit the reliability of hepatic steatosis
393 diagnosis from ultrasound measurements. Xia MF and colleagues
394 proposed an improved method for comparing MRS with standardized
395 US-HRI [23]. In report, the authors tested the contribution of the
396 standardization of the US-HRI using two types of US equipment, and
397 reported high correlation coefficient between US-HRI and MR
398 spectroscopy results. This technique is attractive and promising, but they
399 only tested the standardization approach on two types of US equipment
400 supplied by the same company (GE Healthcare). This makes it difficult to
401 generalize this approach to US units from other suppliers, considering
402 the differences in hardware and postprocessing procedures.

403 There are also several limitations for US-HRI and MRI-PDFF
404 measurements in this study. First, the ROI was limited in size. In
405 subjects with inhomogeneous liver fat distribution, even if multiple ROIs
406 were averaged, there would be no guarantee that the fat content of the
407 entire liver was measured accurately. ROIs were selected manually so
408 the fat evaluation could never be entirely random or entirely objective.
409 Second, the current research employed only one US machine. It is quite
410 possible that results would vary among US units. Moreover, the

411 discrepancy in post-processing algorithms in ultrasound and MRI
412 scanners may limit the correlation between US and MRS. Third, using no
413 histopathology for diagnosing hepatic steatosis as the reference
414 standard may be a potential limitation due to the true prevalence of
415 steatosis not being known with certainty among the participants of the
416 present study. Finally, the measurement of the fat content in MR imaging
417 was based on an available software, and the parameters were not
418 changed from the manufacturer's settings. There was no comparison of
419 parameters to optimize assessment. Additionally, the noise performance
420 was also not examined, leading to the SNR-effect being ignored on the
421 image reconstruction.

422

423 **Conclusions**

424 In conclusion, with MRS-PDFF $\geq 5.56\%$ defined as the gold
425 standard of fatty liver disease, AUCs, cut-off values, sensitivities and
426 specificities of US-HRI and MRI-PDFF were 0.74, 2.75%, 50%, 91.7%
427 and 0.99, 1.54, 100%, 88.9%, respectively. The intraclass correlation
428 coefficients (ICCs) of MRI-PDFF were excellent (0.85), compared to US-
429 HRI (0.70). Therefore, MRI-PDFF was a more reliable technique to for
430 the diagnosis of hepatic steatosis.

431

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442

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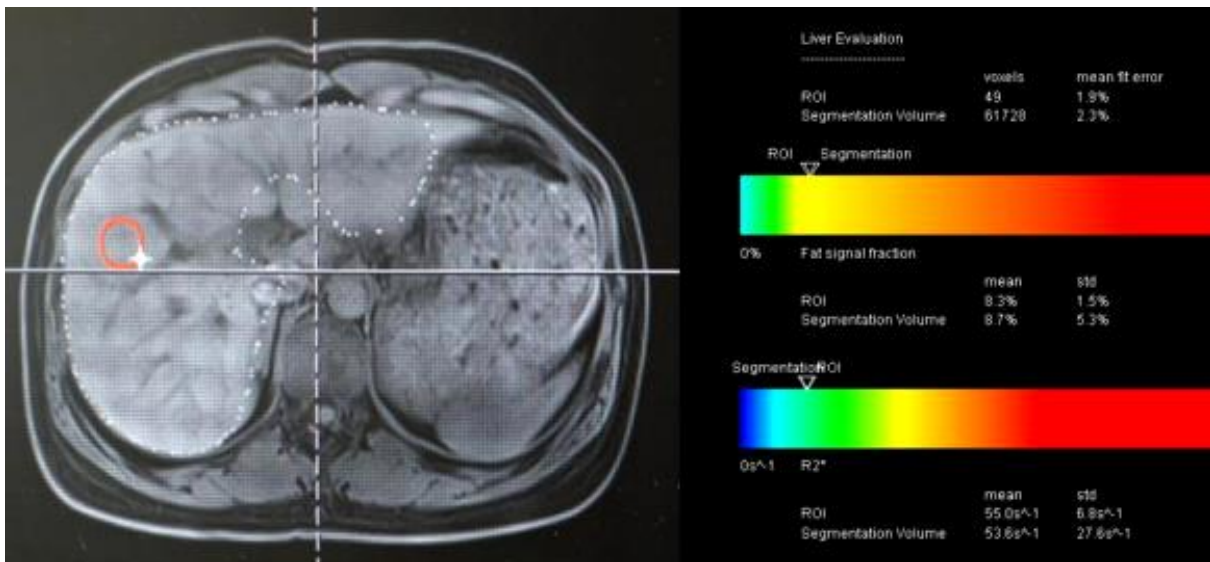
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532

533 Fig 1. HRI measurement on a volunteer with mild hepatic steatosis (HRI = 2.33).

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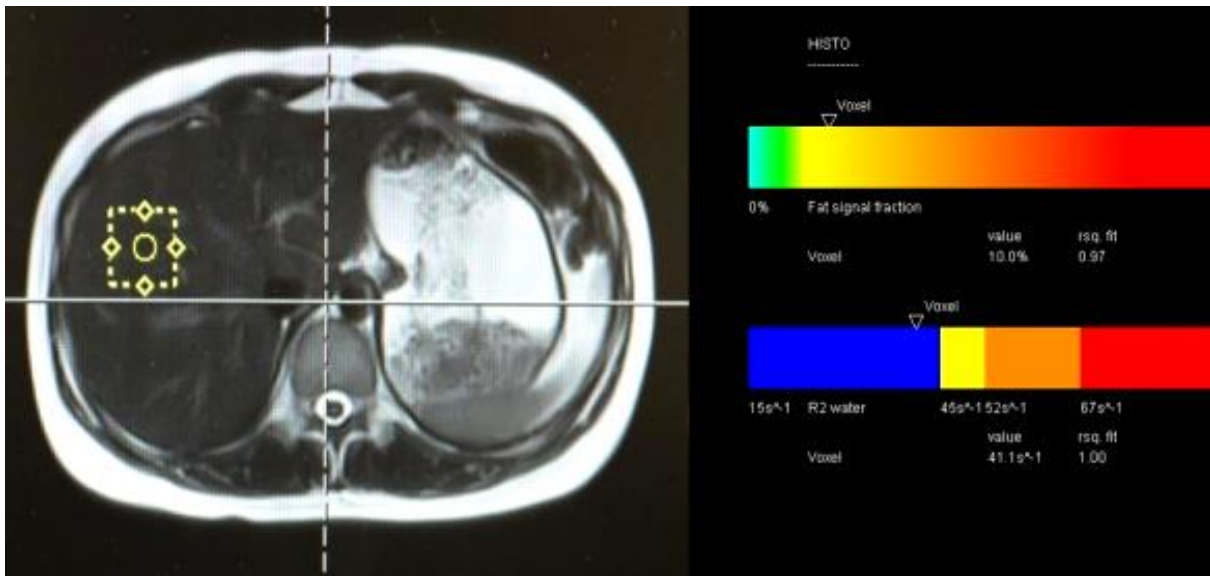


535

536 Fig 2. MRI-PDFF measurement on a volunteer with mild hepatic steatosis (MRI-

537 PDFF = 8.3%).

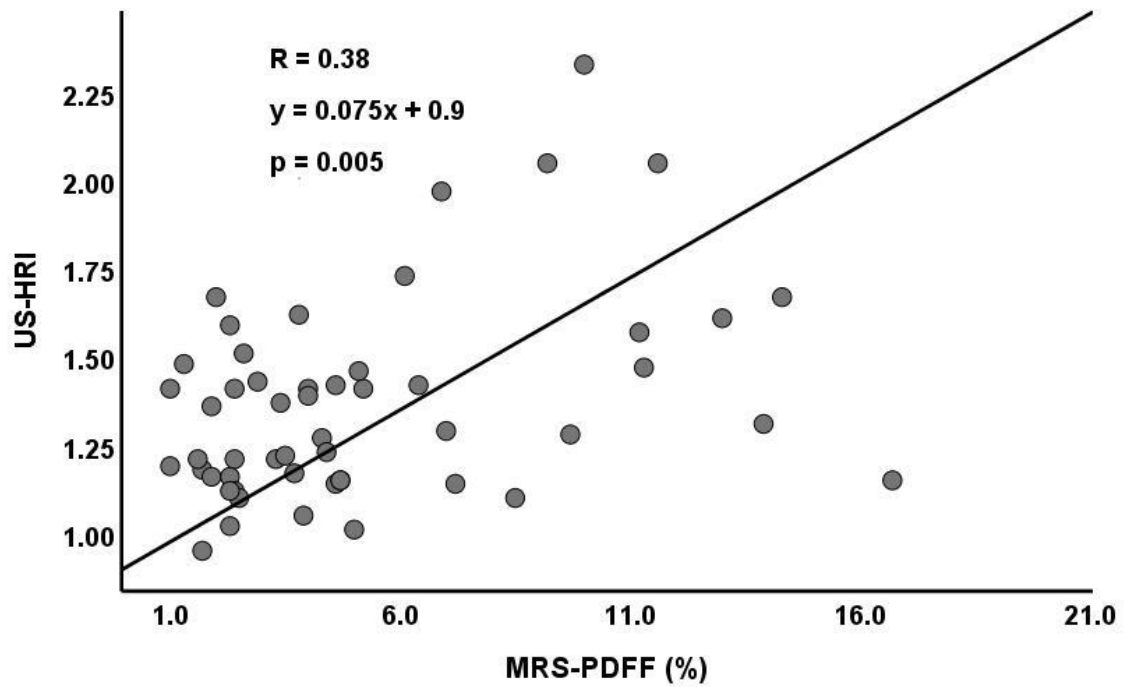
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539

540 **Fig 3. MRS-PDFF measurement on a volunteer with mild hepatic steatosis**
 541 **(MRS-PDFF = 10%).**

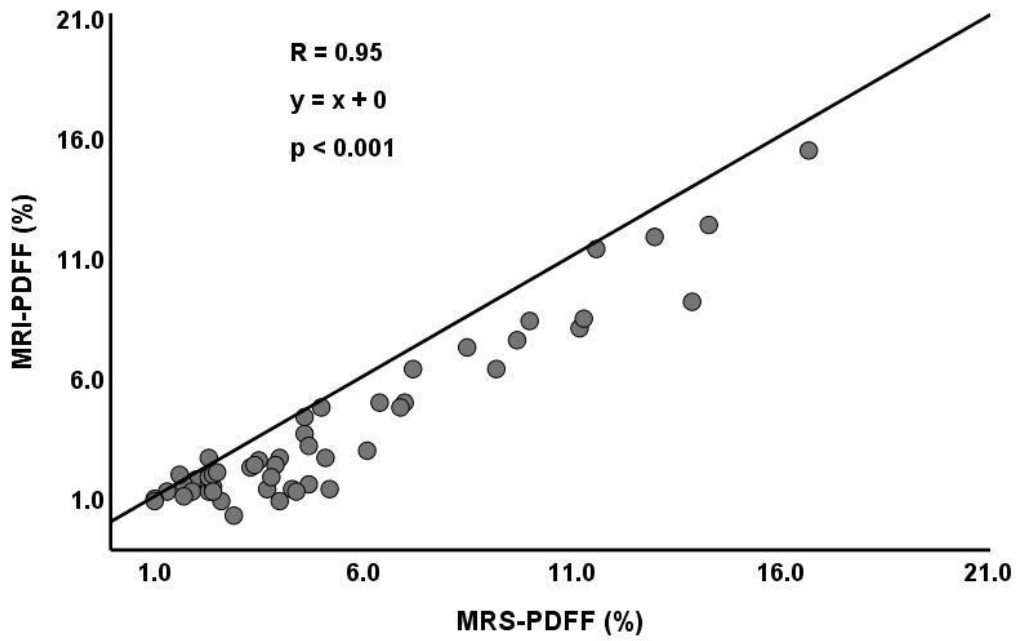
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543

544 **Fig 4. Correlation between US-HRI and MRS-PDFF.**

545

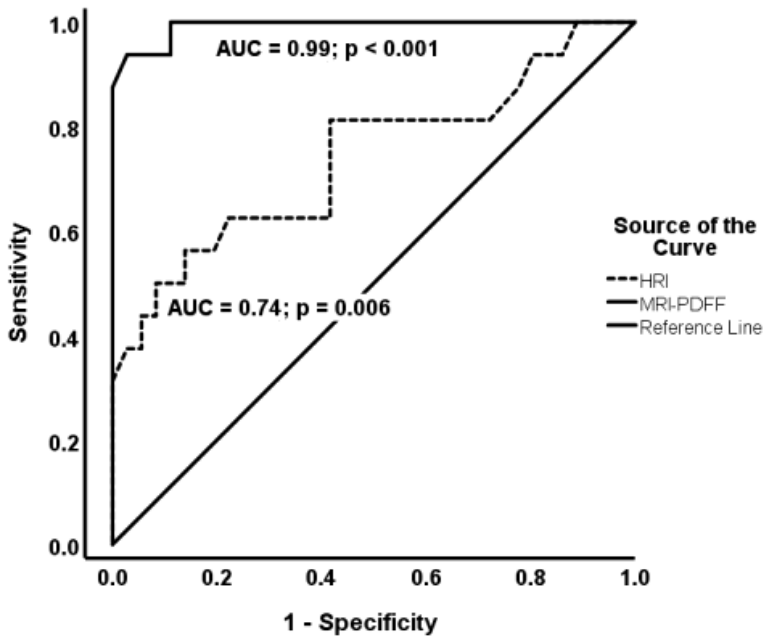


546

547 **Fig 5. Correlation between MRI-PDFF and MRS-PDFF.**

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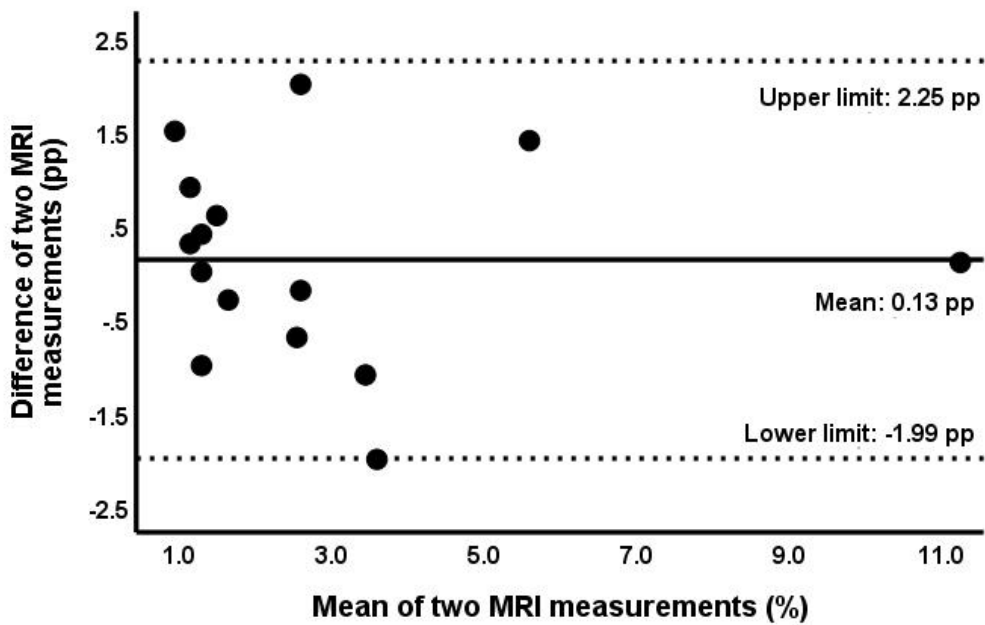
Receiver Operating Characteristics (ROC) Curve



549

550 **Fig 6. ROC curve using the reference of 5.56% as cut-off point in defining**
 551 **diagnostic performance.**

Bland-Altman plots of PDFF measurements generated by using MRI



552

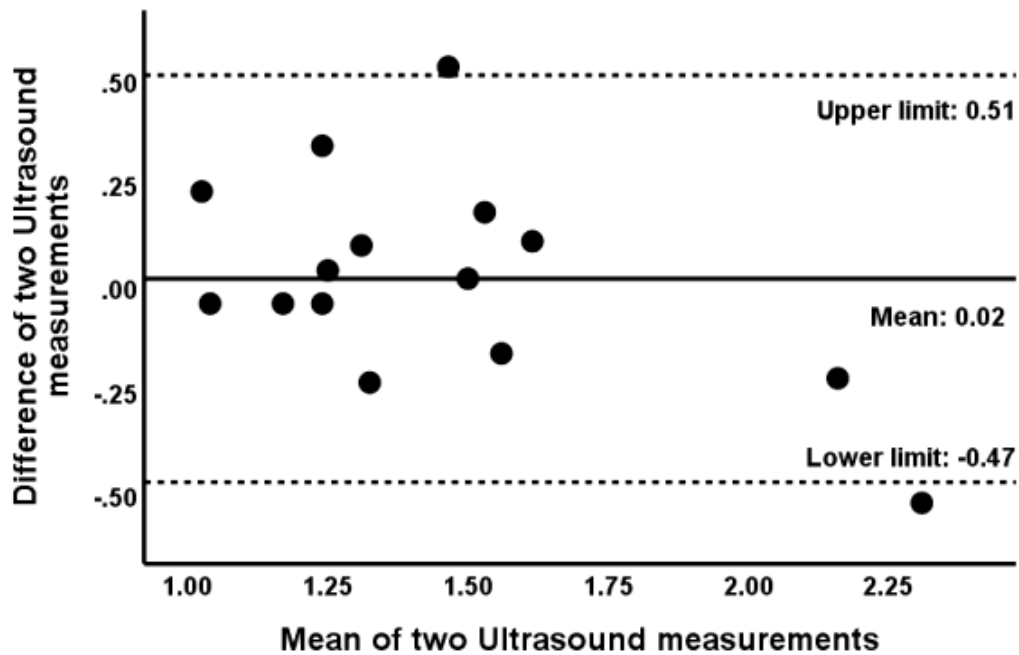
553 **Fig 7. Bland-Altman plots for variability of PDFF measurements generated**
554 **using MRI.**

555 The central line shows the mean of the differences between two PDFF
556 measurements; the dashed lines show upper (mean + 1.96 SD) and lower (mean -
557 1.96 SD) limits of agreement. Here, the mean difference is 0.13 pp, while the limits of
558 agreement are -1.99 pp and 2.25 pp, indicating that 95% of the differences between
559 these two measurements are within this range. The width interval is 4.24 pp.

560

561

Bland-Altman plots of HRI measurements generated by using Ultrasound



562

563 **Fig 8. Bland–Altman plots for variability of HRI measurements generated using**
564 **ultrasound.**

565 The central line shows the mean of the differences between two HRI measurements;
566 the dashed lines show upper (mean + 1.96 SD) and lower (mean - 1.96 SD) limits of
567 agreement. Here, the mean difference is 0.02, while the limits of agreement are -0.47
568 and 0.51, indicating that 95 % of the differences between these two measurements
569 are within this range. The width interval is 0.98.

570